

# OADDL E-News

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## Faculty

**Interim Director:**  
Dr. Jerry Ritchey

**Assistant Director/Quality Manager:**  
Emily J. Cooper

**Microbiology/Molecular  
Diagnostics:**  
Dr. Akhilesh Ramachandran

**Parasitology:**  
Dr. Ruth Scimeca

**Pathology:**  
Dr. Valerie McElliot  
Dr. Craig Miller  
Dr. Sunil Moré  
Dr. Jerry Ritchey  
Dr. Tim Snider

**Serology:**  
Dr. Jerry Ritchey (Interim)

**Graphic Design/Layout:**  
Clarissa Walton

## Robin Madden Receives Stratton Award



**Pictured is Dr. Carlos Risco, Dean, College of Veterinary Medicine and Robin Madden, OADDL Molecular Diagnostics Laboratory Manager.**

Robin Madden, Molecular Diagnostics Laboratory Manager at OADDL, was recently recognized as recipient of the Stratton Staff Award. Established in 1991, the Stratton Staff Award recognizes outstanding service or contributions to the College and represents the highest College-level award for staff recognition.

Ms. Madden is known for being conscientious, dependable and reliable. Robin consistently demonstrates a

high level of expertise in the highly specialized and technical laboratory testing she performs and oversees. Through her collaboration with CVM faculty, she has been instrumental in implementing new PCR tests. She is a well-organized manager who effectively delegates responsibility to expand staff capabilities and productivity. She is committed to achieving her goals, as well as inspiring others to succeed.



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## Thank You ODAFF!!!

OADDL has two exciting projects underway funded by a generous award from the Oklahoma Department of Agriculture, Food & Forestry (\$150,000).

The bulk of the funds will be used to remodel the receiving dock, which will provide a safer and efficient mechanism for the submission of large animals and disposal of animal waste generated through diagnostic testing.

The remainder of the funds were used to purchase two Integra Assist Plus liquid handling units with automated multichannel pipettes. The instruments were placed in the Serology and Molecular Diagnostics Sections. These robotic machines facilitate the processing of large numbers of samples with limited hands on requirements. The automated, hands-free pipetting feature enhances test accuracy while eliminating the potential for laboratory personnel injury from routine repeti-



tive hand movements (such as arthritis or carpal tunnel syndrome).

Both instruments have quickly become essential in high volume, diagnostic testing. In November and December alone, the Serology sec-

tion used the instrument to process approximately 3,900 NPPI surveillance samples and the Molecular Diagnostics section was able to reduce the turnaround time on 1,149 samples submitted during the recent EHV-1 outbreak.

## Important Changes to EIA VS Form 10-11 — Effective Immediately

The VS Form 10-11 is the official Federal EIA test form for EIA test requests. Use of alternative forms must be approved by USDA VS. The VS Form 10-11 was recently updated (FEB2018 VS 10-11 is the current version). **Use of older versions will not be accepted after April 15, 2020.**

Veterinarians completing the VS 10-11 must be a Category II accredited veterinarian authorized to perform accredited duties in the State where the animal was sampled, or a State or Federal animal health official. **Testing laboratories are now required to verify submitting veterinarians are Category II accredited** and must also submit monthly reports of testing performed by state to USDA VS.

All required fields of VS Form 10-11 must be completed and legible. Blank fields are not acceptable.

Results cannot be released until the form is properly completed. The complete Guidance document can be found [here](#).

## Abortion Submission Guidelines

With calving season upon us, January and February are the busiest months for abortions here at OADDL. OADDL provides abortion panels for multiple species that include testing for infectious and toxic (nitrates) causes of abortion in cattle. Although we typically receive individual fetuses, during an outbreak, submission of multiple fetuses (up to 2 for cattle) is encouraged to aid in the diagnosis process. The additional fetus is tested at no additional charge.

The panel includes a necropsy, histopathology (microscopic examination of the tissues), bacterial culture of lungs and abomasal fluids, molecular testing for herpesvirus and BVDV and aqueous fluid testing for nitrates. Microscopic examination of the tissues will help guide selection of other tests if necessary. It is important to submit placenta along with the fetuses, whenever possible, as this is a vital part to our investigation. For a list of the specific tests performed per species, please refer to page 9 of our [Fee Schedule](#).

— Dr. Sierra Salopek



# Sporotrichosis in a Cat and her Veterinarian



Sporotrichosis is a zoonotic fungal infection caused by *Sporothrix schenckii*. Infection is mostly reported in cats and is usually acquired through contaminated wounds. Sporotrichosis in cats poses a significant public health hazard due to the presence of large numbers of organisms in the lesions.

Spread of infection from cats to veterinary professionals is common.

Recently, a skin sample from a cat with a history of non-healing wound on the leg was submitted to OADDL for testing with suspicion of *Bartonella* sp. infection. The cat had scratched the veterinarian and the owner and both of

them developed skin lesions. However, histopathology and fungal culture at OADDL revealed the presence of *Sporothrix schenckii*.

The concerned veterinarian was informed of the results in order to start treatment for sporotrichosis. The clinical features of sporotrichosis in cats include skin lesions with firm, non-painful nodules. Ulceration and fistulation of the nodules with light brown discharge may occur. The lesions are most common on the head and the legs. Regional lymph nodes can be enlarged or fistulated. Autoinoculation to satellite sites is also possible due to grooming.

Histopathology followed by fungal culture is recommended for diagnosis. Deep punch or wedge biopsy specimen from intact non-draining lesions is recommended. Due to its zoonotic potential, personnel handling tissues or animals suspected of sporotrichosis should take necessary biosafety precautions.

— Drs. Sunil Moré & Akhilesh Ramachandran

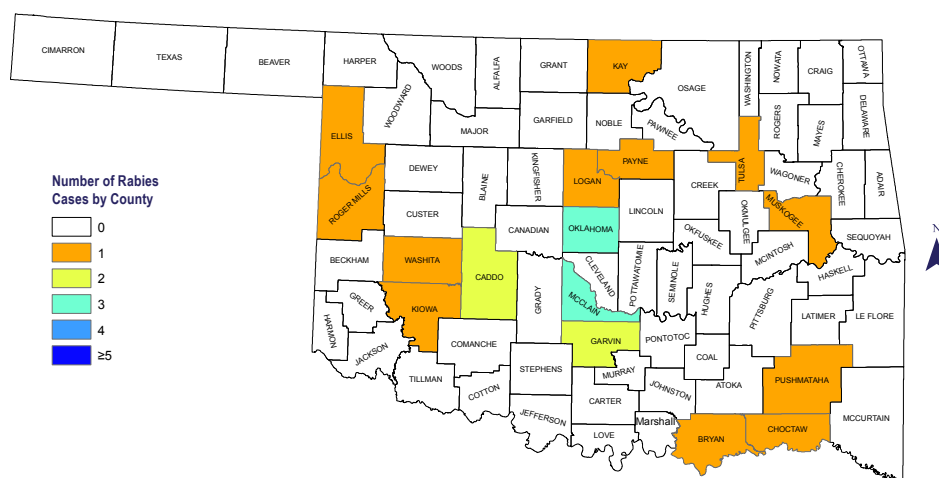
## Changes to Rabies Testing & Submission — Effective Immediately

The daily cutoff time for receipt of specimens for rabies testing at the Oklahoma State Department of Health (OSDH) Public Health Laboratory (PHL) has changed from 11:00 am to 9:00 am. Specimens received after 9:00 am will be tested the following business day.

Rabies specimen submission instructions and an updated *Rabies Specimen Submission* Form can be found [here](#). Reminder, whole animals less than 12 inches long (e.g. bat, small rodent, juvenile animals), whole heads (e.g. dog, cat, skunk, and raccoon) or appropriate brain tissue can be submitted directly to the Rabies Laboratory in Oklahoma City for expedited testing.

If you need assistance with sample collection or have additional diagnostic needs, forward the animal to OADDL for brain removal and necropsy. Standard disposal fees apply for animals bigger than 100 pounds.

### County Location of Animal Rabies in Oklahoma January 1 through November 30, 2019 (N=22)



## Letter from the Interim Director

On behalf of the faculty and staff of OADDL, we wish you a Happy New Year! We appreciate the confidence you have placed in us to handle your animal testing needs. We have been exceptionally busy over November/December with near record accession numbers over what is typically a “quiet” time during the holiday season.

The big news to report is the search process for the new laboratory Director is completed. We are very excited to announce Dr. Jeremiah Saliki has accepted the position. Many of you will remember Dr. Saliki as the Section

Head of Virology/Serology at OADDL, 1993-2005. Dr. Saliki built and directed an outstanding diagnostic program serving clients and livestock producers in the state of Oklahoma. Dr. Saliki is responsible for the production of a monoclonal antibody that is used internationally for BVDV testing and most will not know that he also developed an internationally recognized diagnostic program for marine mammals while here in landlocked Oklahoma.

Since leaving OADDL, Dr. Saliki has served as the Director of the University of Georgia's Diagnostic

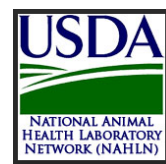
Laboratory (Athens), has been active in the improvement of laboratory management system software and is recognized as an exceptional leader by his peers as he currently serves as the President-elect of the AAVLD.

Dr. Saliki will start April 30. Welcome home Jerry!

Lastly, Dr. Ruth Scimeca has replaced Dr. Yoko Nagamori as our diagnostic parasitologist. Dr. Nagamori has recently accepted a position with Zoetis. We wish her all the best in her new position. Yoko will remain connected with us as an adjunct faculty member. Dr. Scimeca is a veterinarian, holds a PhD degree and is board certified in veterinary parasitology by the American College of Veterinary Microbiologists. More will follow on Dr. Scimeca in the next edition of OADDL E-News.

As always, we appreciate you. Contact us with any questions, comments or concerns.

– Dr. Jerry Ritchey



COME VISIT THE **OADDL** BOOTH AT THE

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VETERINARY  
CONFERENCE

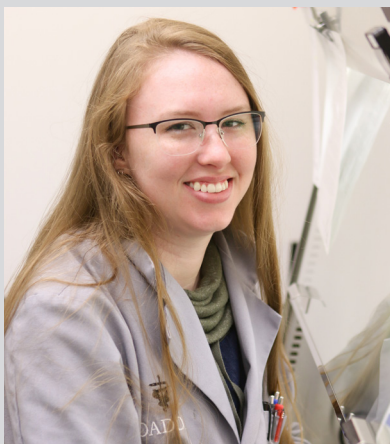
NORMAN, OK  JANUARY, 23-25

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## Getting to Know Us

Kaycee Forsythe joined the Molecular Diagnostics Lab in August 2019 as a Senior Lab Technologist. She received a B.S. in Zoology and a B.S. in Biological Science from Oklahoma State University in 2018. During her weekends, she is a volunteer bottle feeder for Tiny Paws Kitten Rescue. She has one cat, Baguette, and two foster kittens, Pork and Beans.

In her spare time, she enjoys writing, crafting, sewing, and playing with her cats. Ask her about Bryan!



## Ideas/Suggestions for Future Content

We want to hear from you. Send your ideas and suggestions to [oaddl@okstate.edu](mailto:oaddl@okstate.edu).

## Contact Us

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